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SYNTHESIS OF THE 3'-DERIVATIVES OF PHOSPHOROTHIOATE OLIGONUCLEOTIDE ANALOGUES.

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Abstract. 3'-Derivatives of phosphorothioate (PS) oligonucleotide analogues have been synthesized by a selective activation of a 3'-terminal phosphate group of the deprotected PS oligonucleotides using a mixture of triphenylphosphine and 2,2'- dipyridyldisulfide.

Phosphorothioate (PS) oligonucleotide analogues are extensively studied as effective therapeutic agents in the antisense approach¹⁻⁶ In order to enhance an antisense effect, reactive, complementary complex stabilizing and other groups have to be attached to PS oligonucleotides. The well-known methods of preparation of the PS oligonucleotide derivatives on polymer supports ⁴⁻⁶ do not allow to introduce residues bearing active chemical groups into PS olgonucleotides. In these cases, the reactive residues may be deactivated during the deprotection of the support-bond blocked PS oligonucleotides.

The present paper describes a simple and effective method of the synthesis of the various 3'- PS oligonucleotide derivatives (1-7). A selective activation of the 3'-terminal phosphate group of the deblocked PS oligonucleotides with triphenylphosphine (Ph₃P) and 2,2'-dipyridyldisulfide ((PyS)₂) mixture in conditions where the internucleotide phosphorothioate groups are not damaged, is essential to this approach. The method was carried out by using di- and trinucleotide phosphorothioates TpsTp (1a) and TpsTpsTp (1b), bearing a terminal 3'-phosphate group. All obtained derivatives of PS oligonucleotides were detected by ³¹P-NMR-spectroscopy.

The starting phosphorotioates 1a and 1b containing a 3'-terminal phosphate group, were prepared via a phosphotriester approach with using a mixture of N-methylimidazole and 2,4,6-triisopropylbenzenesulfonyl chloride as a condensing reagent in pyridine with deblocking of the protected groups according to Scheme 1. The yields of phosphorothioates 1a and 1b, isolated by the ion exchange chromatography, were 60 and 40%, respectively.

The triphenylphosphine and 2,2'-dipyridyldisulfide mixture was used to activate the terminal phosphate group of di- (1a) and tri- (1b) nucleotide phosphorothioates in the presence of 4-N,N-dimethylaminopyridine (DMAP) or N-methylimidazole (MeIm) as nucleophilic catalysts⁷ (Scheme 2). The reaction time was 5-10 minutes. The yields of the 3'-zwitter-ionic derivatives 2a,b and 3a,b, obtained and isolated by precipitation with ethyl ether, were about 90%.

The prepared zwitter-ionic DMAP (2) or MeIm (3) derivatives were used as intermediates for the synthesis of the 3'-phosphoramidate derivatives of 1a and 1b, bearing an alkylamino group (4), the benzylamine (5) or alkylating N-methyl-4-(N-2-chloroethyl-N-methylamino)benzylamine (6) residues (Scheme 2). The complete disappearance of the ³¹P-NMR resonance signals of DMAP (in area -7 ppm) or MeIm (-11 ppm) derivatives, and the appearance of signals in the area of 6-8 ppm corresponding to phosphoramidate derivatives 4-6, were detected within 5-10 min after addition of

Scheme 1

$$1 + Ph_3P + (PyS)_2 + Nu \longrightarrow 2 \text{ or } 3 \xrightarrow{HR_1} 4.5 \text{ or } 6$$

Nu = N-methylimidazole or 4-N,N-dimethylaminopyridine $Ph_3P = T$ riphenylphosphine $(PyS)_2 = 2,2'$ -dipyridyldisylfide

Scheme 2

10 equivalents of the corresponding amine compounds to the reaction mixtures containing DMAP or MeIm derivatives (0.01 M solution in DMF). The yields of the products, obtained and isolated by precipitation with ethyl ether, were 85-95%.

The N-(2-hydroxyethyl)phenazinium (Phn) derivative **7b** of the PS oligonucleotide was prepared by reaction of the PS olgonucleotide derivative **4**, bearing primary amino group, with Phn⁺Cl⁻ as published in 8. ³¹P-NMR-signals of Phn derivative **7b** in H₂O were registered in area of 55,3 and 55,4 ppm for ps- and 8,7 ppm for terminal phosphoroamidate groups. The yield of **7b** isolated by the ion exchange chromatography, was 85%.

The method proposed allows to introduce different chemical groups, including high reactive (alkylating) ones, at the 3'-end of PS oligonucleotides without the damage of the internucleotide ps-groups of the PS oligonucleotides and formation of any by-products.

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